

Study of complexes between a snake venom or human secreted phospholipase A₂ and calmodulin by high-resolution NMR

Lidija Kovačič¹, Gregor Ilc², Hans Wieng³, Janez Plavec^{2,4,5}, Rolf Boelens³ and Igor Križaj^{1,4,6}

¹ Department of Molecular and Biomedical Sciences, Jožef Stefan Institute, Ljubljana, Slovenia

² Slovenian NMR Center, National Institute of Chemistry, Ljubljana, Slovenia

³ NMR Spectroscopy Research Group, Bijvoet Center for Biomolecular Research, Utrecht University, Utrecht, The Netherlands.

⁴ Faculty of Chemistry and Chemical Technology, University of Ljubljana, Ljubljana, Slovenia

⁵ EN-FIST Center of Excellence, Ljubljana, Slovenia

⁶ Centre of Excellence for Integrated Approaches in Chemistry and Biology of Proteins, Ljubljana, Slovenia

Presenters email address: lidija.kovacic@ijs.si

Secreted phospholipases A₂ (sPLA₂s) are phospholipid-hydrolyzing enzymes, which enzymatic activity is implicated in numerous pathophysiological settings, for example innate immunity, atherosclerosis, cancer, pain, ischemia and Alzheimer's disease. Ammodytoxin (Atx), a neurotoxic sPLA₂ from the *Vipera a. ammodytes* venom, belongs to the group IIA of sPLA₂s and as such presents a model for studying the above motioned pathophysiological processes. Atx forms a high-affinity complex with cytosolic regulatory protein calmodulin (CaM), which results in substantial increase of the enzymatic activity of Atx¹. To understand the role of CaM in the regulation of intra-cytosolic enzyme activity of sPLA₂s, we are currently studying the structure of the complexes formed between Atx or the human group X (hGX) sPLA₂ and CaM using high-resolution NMR spectroscopy. To this end we prepared fully functional recombinant unlabelled and ¹³C/¹⁵N-labelled samples of CaM, Atx and the hGX sPLA₂. ¹⁵N-HSQC spectra of CaM alone and in the complex with unlabelled Atx or hGX sPLA₂ were well resolved, while the ¹⁵N-HSQC spectra of both sPLA₂ alone and in the complex with unlabelled CaM were not, therefore optimization of NMR conditions are still required. The comparison of the spectra of CaM alone and in the complex with either of sPLA₂s revealed several significant changes of chemical shifts in CaM. This can be explained by conformational change of CaM upon its binding to sPLA₂, which seems to be different as reported in cases of CaM binding to other binding proteins. Therefore it seems that sPLA₂s bind to CaM in a unique binding fashion.

Reference:

1. L. Kovačič, M. Novinec, T. Petan, A. Baici, I. Križaj, *Biochemistry* **2009**, 48, 11319–11328.

Acknowledgements: Slovenian Research Agency (P1-0207 and Z1-4071) and Bio-NMR FP7 grants.